#### 1

# **BIOT M02D: BIOPROCESSING: RECOVERY AND PURIFICATION**

#### Originator

achen

#### College

Moorpark College

#### **Attach Support Documentation (as needed)**

BIOT Labor Market Information 032718.docx Biotech LMI data South Central Region 2017-22.xlsx Biotech Job Postings South Central Region Sept 2017- Aug 2018.xlsx

#### Discipline (CB01A)

**BIOT** - Biotechnology

#### Course Number (CB01B)

M02D

#### Course Title (CB02)

Bioprocessing: Recovery and Purification

#### **Banner/Short Title**

Bioprocess:Recovery & Purif

#### **Credit Type**

Credit

#### **Start Term**

Fall 2021

# Co-listed (Same-as) Course(s)

BIOL M12D

#### Taxonomy of Programs (TOP) Code (CB03)

0430.00 - \*Biotechnology and Biomedical Technology

#### SAM Priority Code (CB09)

C - Clearly Occupational

#### **Control Number**

CCC000452381

#### **Primary Minimum Qualification**

**BIOLOGICAL SCIENCES** 

# Department

Biology/Zoology (1021)

#### **Division**

MC EATM, Life & Health Sci

# **Catalog Course Description**

Provides skills training in industrial biotechnology with emphasis on manufacturing pharmaceuticals. Introduces bioprocessing, recovery, and purification techniques. Focuses on protein separation and purification, chromatography, large-scale recovery, and identification of assays. Reviews skills necessary for a successful job search in the field of biotechnology.

# Taxonomy of Programs (TOP) Code (CB03)

0430.00 - \*Biotechnology and Biomedical Technology

#### **Course Credit Status (CB04)**

D (Credit - Degree Applicable)

#### Course Transfer Status (CB05) (select one only)

B (Transferable to CSU only)

#### Course Basic Skills Status (CB08)

N - The Course is Not a Basic Skills Course

#### SAM Priority Code (CB09)

C - Clearly Occupational

#### **Course Cooperative Work Experience Education Status (CB10)**

N - Is Not Part of a Cooperative Work Experience Education Program

#### **Course Classification Status (CB11)**

Y - Credit Course

#### **Educational Assistance Class Instruction (Approved Special Class) (CB13)**

N - The Course is Not an Approved Special Class

# **Course Prior to Transfer Level (CB21)**

Y - Not Applicable

# **Course Noncredit Category (CB22)**

Y - Credit Course

#### **Funding Agency Category (CB23)**

Y - Not Applicable (Funding Not Used)

# Course Program Status (CB24)

1 - Program Applicable

#### **General Education Status (CB25)**

Y - Not Applicable

# **Support Course Status (CB26)**

N - Course is not a support course

#### Field trips

Will not be required

#### **Grading method**

Letter Graded

#### Alternate grading methods

Student Option- Letter/Pass Pass/No Pass Grading

#### Does this course require an instructional materials fee?

No

#### **Repeatable for Credit**

Nο

#### Is this course part of a family?

No

# **Units and Hours**

**Carnegie Unit Override** 

No

#### In-Class

Lecture

**Minimum Contact/In-Class Lecture Hours** 

17.5

**Maximum Contact/In-Class Lecture Hours** 

17.5

**Activity** 

Laboratory

**Minimum Contact/In-Class Laboratory Hours** 

52 5

**Maximum Contact/In-Class Laboratory Hours** 

52.5

# **Total in-Class**

**Total in-Class** 

**Total Minimum Contact/In-Class Hours** 

70

**Total Maximum Contact/In-Class Hours** 

790

# **Outside-of-Class**

Internship/Cooperative Work Experience

**Paid** 

Unpaid

# **Total Outside-of-Class**

**Total Outside-of-Class** 

**Minimum Outside-of-Class Hours** 

35.0

**Maximum Outside-of-Class Hours** 

35.0

# **Total Student Learning**

**Total Student Learning** 

**Total Minimum Student Learning Hours** 

105.0

**Total Maximum Student Learning Hours** 

105.0

**Minimum Units (CB07)** 

^

Maximum Units (CB06)

2

Student Learning Outcomes (CSLOs)				
	Upon satisfactory completion of the course, students will be able to:			
1	use chromatography techniques to successfully purify a candidate protein from a mixture of proteins.			
2	use tangential flow filtration to concentrate a batch of proteins.			
Course Objectives				
	Upon satisfactory completion of the course, students will be able to:			
1	identify and describe protein purification techniques.			
2	demonstrate competency in terminology and acronyms applicable to recovery and purification.			
3	demonstrate skills and knowledge in use of purification equipment.			
4	apply purification concepts to the production of a therapeutic protein.			
5	describe/demonstrate techniques for isolation of proteins from cells and tissue.			
6	identify and explain the function of process equipment.			
7	demonstrate knowledge of and ability to use materials and supplies in purification processes.			

# **Course Content**

#### **Lecture/Course Content**

- 1. (8%) Protein Expression
- 2. (8%) Principles of Protein Separation
- 3. (7%) Recovery-Centrifugation
- 4. (7%) Sterile Filtration
- 5. (7%) Tangential Flow Filtration
- 6. (7%) Affinity Chromatography
- 7. (7%) Ion-exchange Chromatography
- 8. (7%) Principles of Chromatograph Scale-up
- 9. (7%) Contaminants
- 10. (7%) Validation/Scaledown
- 11. (7%)Large Scale Column Packing
- 12. (7%) Immunassay
- 13. (7%) Recovery-Cell Disruption
- 14. (7%) Assay for Identity-SDS-PAGE (Sodium Dodecyl Sulfate Poly-Acrylamide Gel Electrophoresis) Assay

#### **Laboratory or Activity Content**

- 1. (10%) Basic Lab Skills: Lab Notebook, Protocols, Buffer Prep, Protein Concentration
- 2. (30%) Chromatography
- 3. (10%) Protein Analysis
- 4. (20%) Protein Recovery, Folding, and Precipitation
- 5. (20%) Filtration
- 6. (10%) Job Search Preparation

# **Methods of Evaluation**

Which of these methods will students use to demonstrate proficiency in the subject matter of this course? (Check all that apply):

Problem solving exercises Skills demonstrations Written expression

Methods of Evaluation may include, but are not limited to, the following typical classroom assessment techniques/required assignments (check as many as are deemed appropriate):

Classroom Discussion Computational homework Essay exams Group projects Individual projects Journals Laboratory activities Laboratory reports Objective exams Oral presentations **Projects** Problem-solving exams

Participation

Quizzes

Reports/Papers/Journals

Reports/papers

Research papers

Skills demonstrations

# Instructional Methodology

# Specify the methods of instruction that may be employed in this course

Computer-aided presentations Class activities

Class discussions

Case studies

Distance Education

Field trips

Group discussions

Guest speakers

Instructor-guided interpretation and analysis

Instructor-guided use of technology

Internet research

Laboratory activities

Lecture

Practica

#### Describe specific examples of the methods the instructor will use:

- · Discuss the principles of protein separation and show students how to perform protein separation and recovery.
- Show students how to different chromatography techniques (for example, affinity chromatography and ion-exchange chromatography).

# **Representative Course Assignments**

#### Writing Assignments

- 1. Document the purpose, materials and methods, procedure, results, and conclusion for the Tangential Flow Filtration experiment in a lab notebook.
- 2. Write the Standard Operating Procedure (SOP) for purifying a monoclonal antibody from cell culture media using affinity chromatography.
- 3. Keep log books.
- 4. Write reports of experimental results.
- 5. Keep a detailed laboratory notebook.

#### **Critical Thinking Assignments**

- 1. Analyze data obtained from the chromatography column packing experiment to calculate the Height Equivalent of a Theoretical Plate (HETP).
- 2. Analyze chromatograms obtained from chromatography experiments.
- 3. Compose a written summary of experiments and data analysis.
- 4. Solve problem sets.

#### Reading Assignments

- 1. Read papers on protein recovery and purification from Bioprocess International magazine.
- 2. Read the Standard Operating Procedure (SOP) for tangential flow filtration prior to performing the lab.
- 3. Read current biotechnology papers on bioprocessing, recovery, and purification techniques.

#### **Skills Demonstrations**

- 1. Demonstrate chromatography technique at small scale.
- 2. Demonstrate skill in analyzing proteins.

# **Outside Assignments**

# **Representative Outside Assignments**

- 1. Complete problem sets involving calculation of protein yield and purity from purification processes.
- 2. Prepare a group presentation on the process for purifying a specific therapeutic drug product (e.g. Epogen manufactured by Amgen).
- 3. Apply and utilize the CFRs (Code of Federal Regulations).
- 4. Read assigned writings and industry-specific articles.

#### **Articulation**

# **C-ID Descriptor Number**

BIOT 220BX

#### **Status**

**Approved** 

#### **Equivalent Courses at other CCCs**

College	Course ID	Course Title	Units
Contra Costa College	BIOSC 186	Protein Purification and Analysis	2
Irvine Valley College	BIOT 274	Biotechnology B: Proteins	4
Santa Ana College	BIOL 192	Biotech B: Proteins	1.5
Attach Syllabus M02D S19 Course Syllabus.pdf			

# **District General Education**

- **A. Natural Sciences**
- **B. Social and Behavioral Sciences**
- C. Humanities
- D. Language and Rationality
- E. Health and Physical Education/Kinesiology
- F. Ethnic Studies/Gender Studies

#### Course is CSU transferable

Yes

#### **CSU Baccalaureate List effective term:**

S'2008

**CSU GE-Breadth** 

**Area A: English Language Communication and Critical Thinking** 

**Area B: Scientific Inquiry and Quantitative Reasoning** 

**Area C: Arts and Humanities** 

**Area D: Social Sciences** 

**Area E: Lifelong Learning and Self-Development** 

**Area F: Ethnic Studies** 

**CSU Graduation Requirement in U.S. History, Constitution and American Ideals:** 

**IGETC** 

**Area 1: English Communication** 

**Area 2A: Mathematical Concepts & Quantitative Reasoning** 

**Area 3: Arts and Humanities** 

**Area 4: Social and Behavioral Sciences** 

**Area 5: Physical and Biological Sciences** 

**Area 6: Languages Other than English (LOTE)** 

# **Textbooks and Lab Manuals**

# **Resource Type**

Textbook

#### Description

Moorpark College and Industry Partners. Industrial Biotechnology: A Training Manual. Cengage Learning, 2001.

#### **Resource Type**

Textbook

#### Description

Flickinger, Michael, ed. Downstream Industrial Biotechnology: Recovery and Purification. Wiley, 2013.

# **Library Resources**

#### Assignments requiring library resources

Using the Library's print and online resources to locate industry-related articles and to research topics for oral presentations.

#### **Sufficient Library Resources exist**

Yes

# **Example of Assignments Requiring Library Resources**

Use the online library resources to research single-use filtration systems for concentrating protein solutions.

# **Distance Education Addendum**

# **Definitions**

#### **Distance Education Modalities**

Hybrid (51%-99% online) Hybrid (1%-50% online) 100% online

# **Faculty Certifications**

Faculty assigned to teach Hybrid or Fully Online sections of this course will receive training in how to satisfy the Federal and state regulations governing regular effective/substantive contact for distance education. The training will include common elements in the district-supported learning management system (LMS), online teaching methods, regular effective/substantive contact, and best practices.

Yes

Faculty assigned to teach Hybrid or Fully Online sections of this course will meet with the EAC Alternate Media Specialist to ensure that the course content meets the required Federal and state accessibility standards for access by students with disabilities. Common areas for discussion include accessibility of PDF files, images, captioning of videos, Power Point presentations, math and scientific notation, and ensuring the use of style mark-up in Word documents.

Yes

# **Regular Effective/Substantive Contact**

Hybrid (1%-50% online) Modality:

Method of Instruction	Document typical activities or assignments for each method of instruction			
Asynchronous Dialog (e.g., discussion board)	Regular Asynchronous discussion boards will be used to encourage discussion among students where they can compare and contrast/discuss /identify and analyze elements of course outcomes. Other Discussion boards will also be used for Q&A and general class discussion by students and instructor to facilitate student practice in application of recovery and purification in bioprocessing.			
E-mail	Questions and answers between students and instructor, class announcements and memos, assignment submissions by students			
Other DE (e.g., recorded lectures)	Faculty will use a variety of tools and media integrated within the LMS to help students reach SLO such as: o Recorded Lectures, Narrated Slides, Screencasts o Instructor created content o MC Online Library Resources o Canvas Student Groups (Assignments, Discussions) o Websites and Blogs o Multimedia (YouTube, Films on Demand, 3CMedia, Khan Academy, etc.)			
Synchronous Dialog (e.g., online chat)	Synchronous online lectures, online chat forums			
Video Conferencing	Video-conferenced lectures, discussion forums, office hours			
Hybrid (51%-99% online) Modality:				
Method of Instruction	Document typical activities or assignments for each method of instruction			
Asynchronous Dialog (e.g., discussion board)	Regular Asynchronous discussion boards will be used to encourage discussion among students where they can compare and contrast/discuss /identify and analyze elements of course outcomes. Other Discussion boards will also be used for Q&A and general class discussion by students and instructor to facilitate student practice in application of recovery and purification in bioprocessing.			
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Synchronous Dialog (e.g., online chat)	Synchronous online lectures, online chat forums			
Video Conferencing	Video-conferenced lectures, discussion forums, office hours			
100% online Modality:				
Method of Instruction	Document typical activities or assignments for each method of instruction			
Asynchronous Dialog (e.g., discussion board)	Regular Asynchronous discussion boards will be used to encourage discussion among students where they can compare and contrast/discuss /identify and analyze elements of course outcomes. Other Discussion boards will also be used for Q&A and general class discussion by students and instructor to facilitate student practice in application of recovery and purification in bioprocessing.			
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Synchronous Dialog (e.g., online chat)	Synchronous online lectures, online chat forums			
Video Conferencing	Video-conferenced lectures, discussion forums, office hours			
Examinations				
Hybrid (1%-50% online) Modality Online On campus				
Hybrid (51%–99% online) Modality Online On campus				

# **Primary Minimum Qualification**

BIOTECHNOLOGY

# **Review and Approval Dates**

# **Department Chair**

MM/DD/YYYY

# Dean

MM/DD/YYYY

# **Technical Review**

MM/DD/YYYY

# **Curriculum Committee**

10/06/2020

DTRW-I

MM/DD/YYYY

**Curriculum Committee** 

MM/DD/YYYY

**Board** 

01/19/2021

CCCCO

MM/DD/YYYY

DOE/accreditation approval date

MM/DD/YYYY